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Naturally occurring horizontal gene transfer and homologous recombination in *Mycobacterium*

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Acquisition of genetic information through horizontal gene transfer (HGT) is an important evolutionary process by which micro-organisms gain novel phenotypic characteristics. In pathogenic bacteria, for example, it facilitates maintenance and enhancement of virulence and spread of drug resistance. In the genus *Mycobacterium*, to which several primary human pathogens belong, HGT has not been clearly demonstrated. The few existing reports suggesting this process are based on circumstantial evidence of similarity of sequences found in distantly related species. Here, direct evidence of HGT between strains of *Mycobacterium avium* representing two different serotypes is presented. Conflicting evolutionary histories of genes encoding elements of the glycopeptidolipid (GPL) biosynthesis pathway led to an analysis of the GPL cluster genomic sequences from four *Mycobacterium avium* strains. The sequence of *M. avium* strain 2151 appeared to be a mosaic consisting of three regions having alternating identities to either *M. avium* strains 724 or 104. Maximum-likelihood estimation of two breakpoints allowed a ~4100 bp region horizontally transferred into the strain 2151 genome to be pinpointed with confidence. The maintenance of sequence continuity at both breakpoints and the lack of insertional elements at these sites strongly suggest that the integration of foreign DNA occurred by homologous recombination. To our knowledge, this is the first report to demonstrate naturally occurring homologous recombination in *Mycobacterium*. This previously undiscovered mechanism of genetic exchange may have major implications for the understanding of *Mycobacterium* pathogenesis.

INTRODUCTION

Increase in genic content is an important evolutionary mechanism shaping prokaryotic genomes. This dynamic process, counterbalanced by DNA loss, is affected by either duplication of sequences existing in the genome or acquisition of foreign genetic material through horizontal gene transfer (HGT). Novel DNA transferred into the cell is thought to be a major source of new phenotypic characteristics, often allowing survival under strong selection regimes. In pathogenic bacteria, for example, well known cases of transfer include spread of genes conferring antibiotic resistance and virulence determinants (Morschhauser *et al.*, 2000; Ochman *et al.*, 2000; Weigel *et al.*, 2003). HGT can bring not only entirely new sequence families into the genome, but also sequences that are homologous to existing genes (Ochman, 2001), which may lead to a replacement of autochthonous sequences with the acquired copies through homologous recombination.

The genus *Mycobacterium* contains over 70 species of human and animal pathogens as well as non-pathogenic saprophytes. The major human pathogens are *Mycobacterium tuberculosis*, the causative agent of tuberculosis, *Mycobacterium leprae*, causing leprosy, and *Mycobacterium avium*, responsible for opportunistic infections, particularly in AIDS patients. It is estimated that *M. tuberculosis* infects up to one-third of the world's population and results in 3 million deaths annually (Snider, 1994).

In *Mycobacterium* HGT has not been clearly demonstrated. Several existing reports suggesting this process are based on circumstantial evidence involving similarity in sequences found in distantly related bacterial or even eukaryotic species (Gamieldien *et al.*, 2002; Kinsella *et al.*, 2003; Le Dantec *et al.*, 2001; Poelarends *et al.*, 2000). In addition, population genetic studies indicate that, unlike other bacteria, mycobacteria seem not to exchange genetic material between individuals. Genome-wide multilocus analyses of *M. tuberculosis* and *Mycobacterium bovis* environmental samples consistently detected highly significant linkage disequilibrium, suggesting either extreme rarity or non-existence of recombination in natural settings

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Abbreviations: GPL, glycopeptidolipid; HGT, horizontal gene transfer.

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(Smith *et al.*, 2003; Supply *et al.*, 2003). This notion is consistent with early unsuccessful attempts at mutagenesis via allelic exchange (homologous recombination of native gene with an inactivated copy) in these slow-growing mycobacteria. It has been suggested that the unusual structure of the *Mycobacterium recA* gene, which encodes a key protein involved in recombination, DNA repair and regulation of the SOS response (Walker, 1984), may be responsible for inefficient homologous recombination in the *M. tuberculosis* complex (McFadden, 1996). The *recA* gene in these mycobacteria and in *M. leprae* is interrupted by an in-frame ORF encoding an intein that is removed from a precursor protein by a protein-splicing reaction (Davis *et al.*, 1992, 1991). However, subsequent experiments showed that the intein does not affect RecA protein function and the frequency of double cross-over homologous recombination events (Papavinasasundaram *et al.*, 1998). To date, gene knock-outs using allelic exchange have been successfully achieved in both fast-growing *Mycobacterium smegmatis* (Husson *et al.*, 1990) and slow-growing *M. tuberculosis* (Balasubramanian *et al.*, 1996) and *M. bovis* BCG (Aldovini *et al.*, 1993). Pavelka & Jacobs (1999) demonstrated that the recombination frequencies in *M. smegmatis*, *M. tuberculosis* and *M. bovis* were similar, suggesting that the homologous recombination machinery in fast- and slow-growing mycobacteria functions with comparable efficiency.

In another study (Krzywinska *et al.*, 2004) we performed a phylogenetic analysis of 13 *M. avium* strains using as markers two DNA fragments (a 926 bp fragment of the *gtfB* gene and a 2150 bp region spanning *rtfA-mtfC* genes; cf. Fig. 1a) lying in each other's proximity within the gene cluster responsible for biosynthesis of glycopeptidolipids (GPLs). Trees inferred from each marker had significantly incongruent topologies due to a well supported alternative placement of *M. avium* strain 2151. Such incongruence strongly suggests HGT involving one of the markers between the studied strains (Ragan, 2001). Here we present the results of further analysis of the available orthologous sequences of the GPL biosynthesis gene cluster from *M. avium* strains 2151, 104, 724 and A5, and show unequivocal evidence for HGT and homologous recombination in *Mycobacterium*.

METHODS

Nucleotide sequences. The nucleotide sequences of the GPL biosynthesis gene cluster were retrieved from GenBank for the following *M. avium* strains: A5 (accession no. AY130970), 724 (AF125999) and 2151 (AF143772). The corresponding sequence from *M. avium* strain 104 was obtained from The Institute for Genomic Research website (www.tigr.org).

Sequence analysis. The sequences were aligned using CLUSTALX (Thompson *et al.*, 1997). The putative recombinant and parental

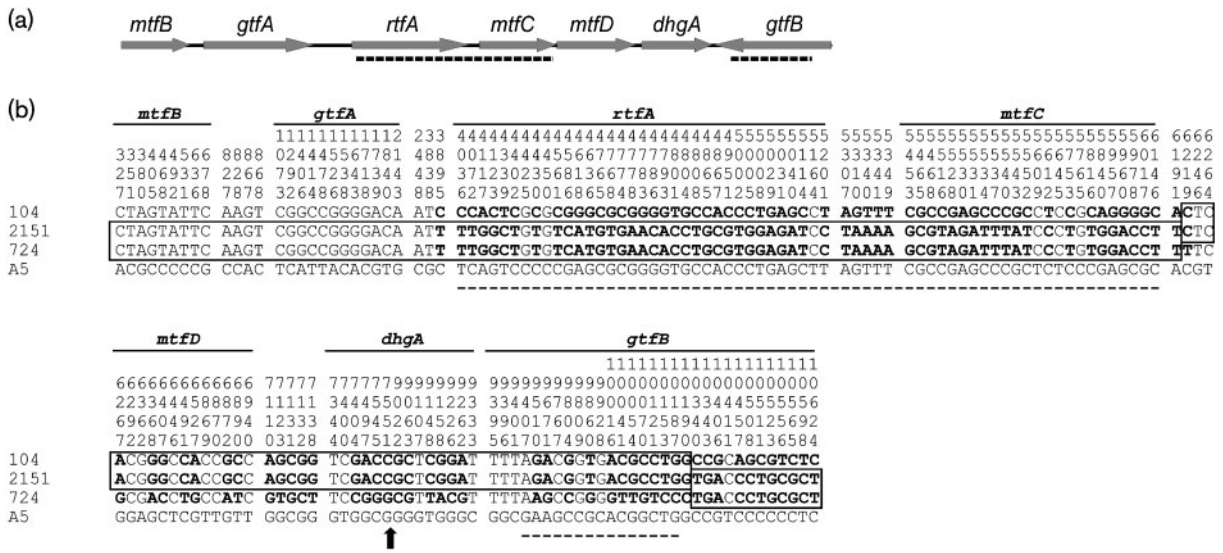

Fig. 1. (a) A schematic of the 5′ end of the GPL biosynthesis cluster in *M. avium* strain A5. Arrows show the position and orientation of individual genes, a line between the arrows represents intergenic spacers and dotted lines under the schematic denote the position of marker sequences yielding discordant phylogenies (Krzywinska *et al.*, 2004). (b). An alignment of the 5′ end of the GPL biosynthesis cluster. The variable positions among four strains of *M. avium* for which the sequence was available are shown; nucleotides at positions variable between strains 104, 2151 and 724 are in bold type. Numbers above the sequences (read from top to bottom) correspond to alignment positions. Portions of identical sequences are boxed; the box around sequences identical in strains 104 and 2151 corresponds to a ~4000 bp fragment horizontally transferred to strain 2151. Contiguous lines above the alignment mark gene sequences and the portions without lines correspond to intergenic spacers. A broken line under the alignment shows the regions corresponding to the marker sequences. The insertion point of the IS*1245* sequence is marked by an arrow.

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sequences were identified upon inspection of variable alignment sites retrieved using MEGA 2.1 (Kumar *et al.*, 2001). The putative recombinant sequence was subjected to a maximum-likelihood analysis using the program LARD developed by Holmes *et al.* (1999). The likelihood of the null hypothesis, H_0, that there has been no recombination event and the likelihood of the hypothesis H_1, that recombination took place, were assessed from a simple unrooted tree of a putative recombinant (strain 2151) and two 'parental' sequences (strains 724 and 104). The likelihood ratio test was performed using the joint likelihood of the two trees on either side of the putative breakpoint and the likelihood of a single tree was calculated on the entire alignment. To assess whether H_1 is a significantly better fit to the data, the likelihood ratio of experimental data was compared to a null distribution obtained by LARD analysis of 500 sequence datasets generated by a Monte Carlo simulation of clonal evolution using Seq-Gen (Rambaut & Grassly, 1997).

RESULTS AND DISCUSSION

The flood of novel sequence information from an increasing number of organisms opens unprecedented opportunities for comparative studies allowing new insights into the fundamental mechanisms of evolutionary change. One of the results of such studies is a growing recognition that HGT is an important evolutionary force acting upon bacterial genomes. However, despite numerous reports in which HGT has been invoked, the actual transfer events are rarely observed (Ochman, 2001).

Our recent phylogenetic analysis of *M. avium* strains using two closely linked markers, *gtfB* and *rtfA-mtfC* from the GPL biosynthesis gene cluster, unexpectedly revealed a potential horizontal transfer of one of the marker sequences between the strains belonging to different serotypes (Krzywinska *et al.*, 2004). The notion of transfer was based on a well-supported alternative position of one of the strains (i.e. 2151) in the trees inferred from each marker individually. Inspection of the sequence substitution patterns from all studied strains revealed that within the *rtfA-mtfC* marker, strain 2151 shared identical sequence with strain 724, whereas within the *gtfB* gene fragment it was identical to five other strains, including 104, but substantially different from strain 724.

To further explore the transfer hypothesis and identify a potential sequence breakpoint, we retrieved and analysed all *M. avium* genomic sequences that exist in the publicly available databases which corresponded to the GPL cluster. These included sequences from strains 104, 724, 2151 and A5. We focused our study on the highly conserved 5′ region of the GPL cluster stretching between the *mtfB* and *gtfB* genes (Krzywinska & Schorey, 2003). The alignment of the four 10 624 bp sequences was unambiguous. The pattern of the variable alignment positions within that region provided clear evidence for a mosaic structure within the strain 2151 genomic sequence (Fig. 1). Its 5′ half, containing the *rtfA-mtfC* marker, was identical to the sequence from strain 724. A potential breakpoint was located within the intergenic spacer between *mtfC* and *mtfD* genes, at which point the subsequent sequence of strain 2151 shared identity with strain 104. Remarkably, another potential breakpoint was identified at the end of the aligned region, within the *gtfB* gene. Downstream from the *gtfB* marker sequence, the pattern of distribution of nucleotide substitutions reverted to the one observed within the 5′ half of the alignment. To substantiate our hypothesis of gene transfer we performed sequence analysis using the LARD maximum-likelihood method (Holmes *et al.*, 1999), in which the likelihoods of the null hypothesis, H_0, (i.e. there has been no recombination event at the analysed regions) and of the alternative hypothesis, H_1, were calculated. This method allowed us to identify the two breakpoint regions within the alignment, which were found to be before positions 6219 and 10340. Monte Carlo simulation used to assess whether H_1 is a significantly better fit to the data (i.e. that the likelihood ratio of H_1 to H_0 is greater than we would expect by chance) revealed that the log likelihood ratio of the real data (recombination event involving region spanning positions 6219–10 340) was significantly greater than for any of the simulated dataset (73·4 vs maximum of 15·09). A mosaic structure of the aligned region with blocks of sequences having unambiguously different evolutionary affiliations and identification of both breakpoints allowed us to pinpoint with confidence an entire region horizontally transferred into strain 2151. The maintenance of sequence continuity (lack of indels) and the absence of mobile elements within both breakpoints clearly points to homologous recombination as a mechanism of the foreign DNA integration into the strain 2151 genome.

It is worth noting that within the recombination region an insertion sequence IS*1245* was embedded, although the significance of this finding remains unknown. Insertion sequence (IS) elements are known to modify a bacterial genome by mediating mobility of DNA fragments within an individual. The widespread occurrence and high degree of similarity between IS elements in different *Mycobacterium* species suggest that they spread across members of the genus through horizontal transfer (Gordon *et al.*, 1999; Howard *et al.*, 2002). However, evidence from the analysis of the *M. tuberculosis* H37Rv genome shows no apparent link between potential IS transfer and introduction of novel genes into the bacillus (Gordon *et al.*, 1999).

Correct identification of ancient HGT events is difficult and is associated with a high degree of uncertainty. Many of the reports claiming such ancient transfer based on sequence similarity or phylogenetic distribution of sequences are likely to be false positives, and at least one such case apparently concerns mycobacteria. It has been proposed that 19 genes found in the *M. tuberculosis* genome, and implicated to have a role in mycobacterial pathogenesis, are of eukaryotic origin (Gamieldien *et al.*, 2002). However, this finding was shown to be an artifact of insufficient taxon sampling, when the genes were reanalysed in the context of a larger dataset (Kinsella & McInerney, 2003). Recently Kinsella *et al.* (2003) suggested that a close phylogenetic affiliation of genes involved in fatty acid

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biosynthesis in mycobacteria and proteobacteria, two distantly related groups of organisms, is a signature of HGT. Although plausible, considering a broad spectrum of taxa used for the analysis, their hypothesis leaves a tinge of suspicion, because the codon usage bias in the putatively transferred genes was typical of *Mycobacterium* genes. For this reason, ancient gene duplications and subsequent retention of sequences in unrelated lineages can serve as an alternative hypothesis explaining the pattern observed in that study.

Apparently, recent gene transfer has been implicated in the evolution of the catabolic pathway for the man-made chemical 1,2-dibromoethane, found in *Mycobacterium* sp. strain GP1 (Poelarends *et al.*, 2000). This highly toxic compound is degraded by the haloalkane dehalogenase (*dhaA*) gene product. A highly conserved *dhaA* gene was also found in phylogenetically distant *Pseudomonas pavonaceae* and *Rhodococcus rhodochrous* (Kulakova *et al.*, 1997), organisms isolated, like *Mycobacterium* sp. strain GP1, from different strongly contaminated environmental samples. Remarkably, in *Rhodococcus rhodochrous* the *dhA* region was located on the autotransmissible plasmid pRTL1 (Kulakova *et al.*, 1997). Another case of potentially recent HGT was suggested by the sequence analysis of the linear plasmid pCLP from *Mycobacterium celatum*, which revealed loci with high nucleotide sequence identity to loci on the *M. tuberculosis* chromosome (Le Dantec *et al.*, 2001). Moreover, the acquisition of a plasmid encoding mycolactone, a key toxin responsible for skin tissue destruction, was likely to be a recent event, resulting in the emergence of *Mycobacterium ulcerans* as a causative agent of Buruli ulcer (Stinear *et al.*, 2004). Whereas the mycolactone gene cluster in *M. ulcerans* remained on a plasmid, the other two examples of presumably recent HGT implied incorporation of the transferred genes into the genome by either excisive-integrative recombination or integrase-dependent acquisition. Our study is the first to report naturally occurring transfer and incorporation by homologous recombination of the transferred DNA into the *Mycobacterium* chromosome. The transfer most likely occurred very recently, since the recombination event has not been obscured by subsequent nucleotide substitutions within the replaced region, as shown by its comparison with the parental sequence. Recent successful allelic exchange experiments confirmed the feasibility of homologous recombination in *M. avium* (Maslow *et al.*, 2003; Otero *et al.*, 2003; Krzywinska & others, unpublished). In two of those studies a gene knock-out has been achieved in the *rtfA* and *mtfD* genes located within the GPL cluster (Maslow *et al.*, 2003; Krzywinska & others, unpublished).

Several mechanisms have been proposed for the exchange of chromosomal DNA between bacteria by homologous recombination as a consequence of conjugation, transduction and transformation. However, there is little information regarding the significance of these mechanisms in *Mycobacterium*. It has been shown that mycobacterial plasmids can replicate within most *Mycobacterium* species, so they can theoretically be spread horizontally, promoting gene transfer between mycobacteria (Kirby *et al.*, 2002; Le Dantec *et al.*, 2001). Several plasmids were found in the *M. avium* complex (MAC) (Hellyer *et al.*, 1991; Meissner & Falkinham, 1986), among them plasmid pVT2, which is thought to be conjugative (Kirby *et al.*, 2002), and raises the possibility for conjugative transfer in MAC – a mechanism known to occur between *M. smegmatis* strains (Parsons *et al.*, 1998) and between *M. smegmatis* and other bacterial species (Gormley & Davies, 1991). The widespread occurrence of related plasmids from *M. avium*, *Mycobacterium intracellulare* and *Mycobacterium scrofulaceum* (Jucker & Falkinham, 1990) suggests that the plasmids have the ability to transfer between hosts in the environment, where MAC strains are ubiquitous. Transfer can also occur via a transformation process. Spontaneous plasmid transformation was discovered in *M. smegmatis* (Bhatt *et al.*, 2002) and natural competence for transformation by chromosomal DNA was reported in *M. avium* (Tsukamura *et al.*, 1960). Moreover, DNA fragments can be transferred between bacterial cells in a transduction process mediated by bacteriophages, over 250 of which have been described from *Mycobacterium* (Hatfull & Jacobs, 1994). Genomic characterization of 14 mycobacteriophages revealed within their genomes over 50 genes not associated with phage growth (Pedulla *et al.*, 2003). Intriguingly, two of these phages contained a close homologue of a gene found in *M. leprae* and *M. tuberculosis*, which encodes the antigen Lsr2, a strong stimulator of the immune response, hinting at a possible role of phages in mycobacterial virulence. It is not clear if mycobacteriophages transmit such genes to bacterial genomes (Pedulla *et al.*, 2003); however, experiments in which mycobacteriophages have been used to create systems for gene delivery in mycobacteria (Bardarov *et al.*, 1997) show the feasibility of such a process.

Within pathogenic species, recombination may have important ramifications regarding adaptation to host, response to control or tracing of strains for epidemiological purposes (Ochman, 2001). The presence of different gene delivery mechanisms and functional homologous recombination machinery in various *Mycobacterium* species raises the possibility of naturally occurring transfer and recombination not only in *M. avium*, but also in other members of the genus. Moreover, the evidence presented in this study may contribute to the understanding of the mechanisms of drug resistance development in *Mycobacterium*. In *M. tuberculosis* all antibiotic resistance is chromosomally mediated and results from point mutations in different genes (depending on the antibiotic), rather than from acquisition of new genetic elements encoding antibiotic-altering enzymes (Musser, 1995). The rate of spontaneous mutations *in vitro* is low in *M. tuberculosis*, relative to the range noticed in other bacteria (David & Newman, 1971). This contrasts with a high frequency of emergence of drug resistance in tuberculosis patients (Espinal *et al.*, 2000). Likewise, multiple drug resistance, which occurs in 1–3 %

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of total tuberculosis isolates, is inexplicably high. It has been shown that DnaE2 polymerase, which is responsible for inducible mutagenesis in *M. tuberculosis*, can contribute directly to the appearance of resistance *in vivo* (Boshoff *et al.*, 2003). Homologous recombination brings a new potential explanation for the generation of multiple drug resistance. Although HGT has not been shown to be involved in the generation of drug resistance, it is possible that the mutated allele conferring resistance to a given antibiotic in one *Mycobacterium* can be transferred to another. If this isolate, containing a different resistance phenotype, incorporated the mutated allele into its genome through homologous recombination, one could obtain a multidrug-resistant strain. Simultaneous infection of patients by two different strains of *M. tuberculosis* (Braden *et al.*, 2001) or *M. avium* (von Reyn *et al.*, 1995) also creates the opportunity for such a DNA exchange.

In summary, we have unequivocally shown naturally occurring homologous recombination within the GPL biosynthesis cluster of *M. avium* 2151. At present, it is not yet known how often recombination of highly selected loci contributes to pathogenesis or the generation of multidrug resistance in mycobacteria.

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REFERENCES

Aldovini, A., Husson, R. N. & Young, R. A. (1993). The *uraA* locus and homologous recombination in *Mycobacterium bovis* BCG. *J Bacteriol* **175**, 7282–7289.

Balasubramanian, V., Pavelka, M. S. J., Bardarov, S. S., Martin, J., Weisbrod, T. R., McAdam, R. A., Bloom, B. R. & Jacobs, W. R., Jr (1996). Allelic exchange in *Mycobacterium tuberculosis* with long linear recombination substrates. *J Bacteriol* **178**, 273–279.

Bardarov, S., Kriakov, J., Carriere, C. & 7 other authors (1997). Conditionally replicating mycobacteriophages: a system for transposon delivery to *Mycobacterium tuberculosis*. *Proc Natl Acad Sci U S A* **94**, 10961–10966.

Bhatt, A., Kieser, H. M., Melton, R. E. & Kieser, T. (2002). Plasmid transfer from *Streptomyces* to *Mycobacterium smegmatis* by spontaneous transformation. *Mol Microbiol* **43**, 135–146.

Boshoff, H. I., Reed, M. B. & Barry, C. E. (2003). DnaE2 polymerase contributes to *in vivo* survival and the emergence of drug resistance in *Mycobacterium tuberculosis*. *Cell* **113**, 183–193.

Braden, C. R., Morlock, G. P., Woodley, C. L. & 7 other authors (2001). Simultaneous infection with multiple strains of *Mycobacterium tuberculosis*. *Clin Infect Dis* **33**, 42–47.

David, H. L. & Newman, C. M. (1971). Some observations on the genetics of isoniazid resistance in the tubercle bacilli. *Am Rev Respir Dis* **104**, 508–515.

Davis, E. O., Sedgwick, S. G. & Colston, M. J. (1991). Novel structure of the *recA* locus of *Mycobacterium tuberculosis* implies processing of the gene product. *J Bacteriol* **173**, 5653–5662.

Davis, E. O., Jenner, P. J., Brooks, P. C., Colston, M. J. & Sedgwick, S. G. (1992). Protein splicing in the maturation of *M. tuberculosis* RecA protein: a mechanism for tolerating a novel class of intervening sequence. *Cell* **71**, 201–210.

Espinal, M. A., Kim, S. J., Suarez, P. G. & 7 other authors (2000). Standard short-course chemotherapy for drug-resistant tuberculosis: treatment outcomes in 6 countries. *JAMA* **283**, 2537–2545.

Gamieldien, J., Ptitsyn, A. & Hide, W. (2002). Eukaryotic genes in *Mycobacterium tuberculosis* could have a role in pathogenesis and immunomodulation. *Trends Genet* **18**, 5–8.

Gordon, S. V., Heym, B., Parkhill, J., Barrell, B. & Cole, S. T. (1999). New insertion sequences and a novel repeated sequence in the genome of *Mycobacterium tuberculosis* H37Rv. *Microbiology* **145**, 881–892.

Gormley, E. P. & Davies, J. (1991). Transfer of plasmid RSF1010 by conjugation from *Escherichia coli* to *Streptomyces lividans* and *Mycobacterium smegmatis*. *J Bacteriol* **173**, 6705–6708.

Hatfull, G. F. & Jacobs, W. R. J. (1994). Mycobacteriophages: cornerstones of mycobacterial research. In *Tuberculosis: Pathogenesis, Protection and Control*, pp. 165–183. Edited by B. R. Bloom. Washington, DC: American Society for Microbiology.

Hellyer, T. J., Brown, I. N., Dale, J. W. & Easmon, C. S. (1991). Plasmid analysis of *Mycobacterium avium-intracellulare* (MAI) isolated in the United Kingdom from patients with and without AIDS. *J Med Microbiol* **34**, 225–231.

Holmes, E. C., Worobey, M. & Rambaut, A. (1999). Phylogenetic evidence for recombination in dengue virus. *Mol Biol Evol* **16**, 405–409.

Howard, S. T., Byrd, T. F. & Lyons, C. R. (2002). A polymorphic region in *Mycobacterium abscessus* contains a novel insertion sequence element. *Microbiology* **148**, 2987–2996.

Husson, R. N., James, B. E. & Young, R. A. (1990). Gene replacement and expression of foreign DNA in mycobacteria. *J Bacteriol* **172**, 519–524.

Jucker, M. T. & Falkinham, J. O. (1990). Epidemiology of infection by nontuberculous mycobacteria IX. Evidence for two DNA homology groups among small plasmids in *Mycobacterium avium*, *Mycobacterium intracellulare*, and *Mycobacterium scrofulaceum*. *Am Rev Respir Dis* **142**, 858–862.

Kinsella, R. J. & McInerney, J. O. (2003). Eukaryotic genes in *Mycobacterium tuberculosis*? Possible alternative explanations. *Trends Genet* **19**, 687–689.

Kinsella, R. J., Fitzpatrick, D. A., Creevey, C. J. & McInerney, J. O. (2003). Fatty acid biosynthesis in *Mycobacterium tuberculosis*: lateral gene transfer, adaptive evolution, and gene duplication. *Proc Natl Acad Sci U S A* **100**, 10320–10325.

Kirby, C., Waring, A., Griffin, T. J. & Falkinham, J. O. (2002). Cryptic plasmids of *Mycobacterium avium*: Tn*552* to the rescue. *Mol Microbiol* **43**, 173–186.

Krzywinska, E. & Schorey, J. S. (2003). Characterization of genetic differences between *Mycobacterium avium* subsp. *avium* strains of diverse virulence with a focus on the glycopeptidolipid biosynthesis cluster. *Vet Microbiol* **91**, 249–264.

Krzywinska, E., Krzywinski, J. & Schorey, J. S. (2004). Phylogeny of *Mycobacterium avium* strains inferred from glycopeptidolipid biosynthesis pathway genes. *Microbiology* **150**, 1699–1706.

Kulakova, A. N., Larkin, M. J. & Kulakov, L. A. (1997). The plasmid-located haloalkane dehalogenase gene from *Rhodococcus rhodochrous* NCIMB13064. *Microbiology* **143**, 109–115.

Kumar, S., Tamura, K., Jakobsen, I. B. & Nei, M. (2001). MEGA2: molecular evolutionary genetics analysis software. *Bioinformatics* **17**, 1244–1245.

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Le Dantec, C., Winter, N., Gicquel, B., Vincent, V. & Picardeau, M. (2001). Genomic sequence and transcriptional analysis of a 23-kilobase mycobacterial linear plasmid: evidence for horizontal transfer and identification of plasmid maintenance systems. *J Bacteriol* **183**, 2157–2164.

Maslow, J. N., Irani, V. R., Lee, S. H., Eckstein, T. M., Inamine, J. M. & Belisle, J. T. (2003). Biosynthetic specificity of the rhamnosyltransferase gene of *Mycobacterium avium* serovar 2 as determined by allelic exchange mutagenesis. *Microbiology* **149**, 3193–3202.

McFadden, J. (1996). Recombination in mycobacteria. *Mol Microbiol* **21**, 205–211.

Meissner, P. S. & Falkinham, J. O., 3rd (1986). Plasmid DNA profiles as epidemiological markers for clinical and environmental isolates of *Mycobacterium avium*, *Mycobacterium intracellulare*, and *Mycobacterium scrofulaceum*. *J Infect Dis* **153**, 325–331.

Morschhauser, J., Kohler, G., Ziebuhr, W., Blum-Oehler, G., Dobrindt, U. & Hacker, J. (2000). Evolution of microbial pathogens. *Philos Trans R Soc Lond B Biol Sci* **355**, 695–704.

Musser, J. M. (1995). Antimicrobial agent resistance in mycobacteria: molecular genetic insights. *Clin Microbiol Rev* **8**, 496–514.

Ochman, H. (2001). Lateral and oblique gene transfer. *Curr Opin Genet Dev* **11**, 616–619.

Ochman, H., Lawrence, J. G. & Groisman, E. A. (2000). Lateral gene transfer and the nature of bacterial innovation. *Nature* **405**, 299–304.

Otero, J., Jacobs, W. R. J. & Glickman, M. S. (2003). Efficient allelic exchange and transposon mutagenesis in *Mycobacterium avium* by specialized transduction. *Appl Environ Microbiol* **69**, 5039–5044.

Papavinasasundaram, K. G., Colston, M. J. & Davis, E. O. (1998). Construction and complementation of a *recA* deletion mutant of *Mycobacterium smegmatis* reveals that the intein in *Mycobacterium tuberculosis recA* does not affect RecA function. *Mol Microbiol* **30**, 525–534.

Parsons, L. M., Jankowski, C. S. & Derbyshire, K. M. (1998). Conjugal transfer of chromosomal DNA in *Mycobacterium smegmatis*. *Mol Microbiol* **28**, 571–582.

Pavelka, M. S. J. & Jacobs, W. R. J. (1999). Comparison of the construction of unmarked deletion mutations in *Mycobacterium smegmatis*, *Mycobacterium bovis* bacillus Calmette-Guerin, and *Mycobacterium tuberculosis* H37Rv by allelic exchange. *J Bacteriol* **181**, 4780–4789.

Pedulla, M. L., Ford, M. E., Houtz, J. M. & 17 other authors (2003). Origins of highly mosaic mycobacteriophage genomes. *Cell* **113**, 171–182.

Poelarends, G. J., Kulakov, L. A., Larkin, M. J., van Hylckama Vlieg, J. E. & Janssen, D. B. (2000). Roles of horizontal gene transfer and gene integration in evolution of 1,3-dichloropropene- and 1,2-dibromoethane-degradative pathways. *J Bacteriol* **182**, 2191–2199.

Ragan, M. A. (2001). Detection of lateral gene transfer among microbial genomes. *Curr Opin Genet Dev* **11**, 620–626.

Rambaut, A. & Grassly, N. C. (1997). Seq-Gen: an application for the Monte Carlo simulation of DNA sequence evolution along phylogenetic trees. *Comput Appl Biosci* **13**, 235–238.

Smith, N. H., Dale, J., Inwald, J., Palmer, S., Gordon, S. V., Hewinson, R. G. & Smith, J. M. (2003). The population structure of *Mycobacterium bovis* in Great Britain: clonal expansion. *Proc Natl Acad Sci U S A* **100**, 15271–15275.

Snider, D. E. J. (1994). Global burden of tuberculosis. In *Tuberculosis: Pathogenesis, Protection, and Control*, pp. 3–59. Edited by B. R. Bloom. Washington DC: American Society for Microbiology.

Stinear, T. P., Mve-Obiang, A., Small, P. L. C. & 12 other authors (2004). Giant plasmid-encoded polyketide synthases produce the macrolide toxin of *Mycobacterium ulcerans*. *Proc Natl Acad Sci U S A* **101**, 1345–1349.

Supply, P., Warren, R. M., Banuls, A. L. & 6 other authors (2003). Linkage disequilibrium between minisatellite loci supports clonal evolution of *Mycobacterium tuberculosis* in a high tuberculosis incidence area. *Mol Microbiol* **47**, 529–538.

Thompson, J. D., Gibson, T. J., Plewniak, F., Jeanmougin, F. & Higgins, D. G. (1997). The CLUSTAL_X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Res* **25**, 4876–4882.

Tsukamura, M., Hashimoto, H. & Noda, Y. (1960). Transformation of isoniazid and streptomycin resistance in *Mycobacterium avium* by desoxyribonucleate derived from isoniazid- and streptomycin-double-resistant cultures. *Am Rev Respir Dis* **81**, 403–406.

von Reyn, C. F., Jacobs, N. J. R., Arbeit, D., Maslow, J. N. & Niemczyk, S. (1995). Polyclonal *Mycobacterium avium* infections in patients with AIDS: variations in antimicrobial susceptibilities of different strains of *M. avium* isolated from the same patient. *J Clin Microbiol* **33**, 1008–1010.

Walker, G. C. (1984). Mutagenesis and inducible responses to deoxyribonucleic acid damage in *Escherichia coli*. *Microbiol Rev* **48**, 60–93.

Weigel, L. M., Clewell, D. B., Gill, S. R. & 7 other authors (2003). Genetic analysis of a high-level vancomycin-resistant isolate of *Staphylococcus aureus*. *Science* **302**, 1569–1571.

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